

Challenges in developing IVD tests for innovative biomarkers: what an IVD manufacturer needs for accurate measurements

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IVDR (EU) 2017/746



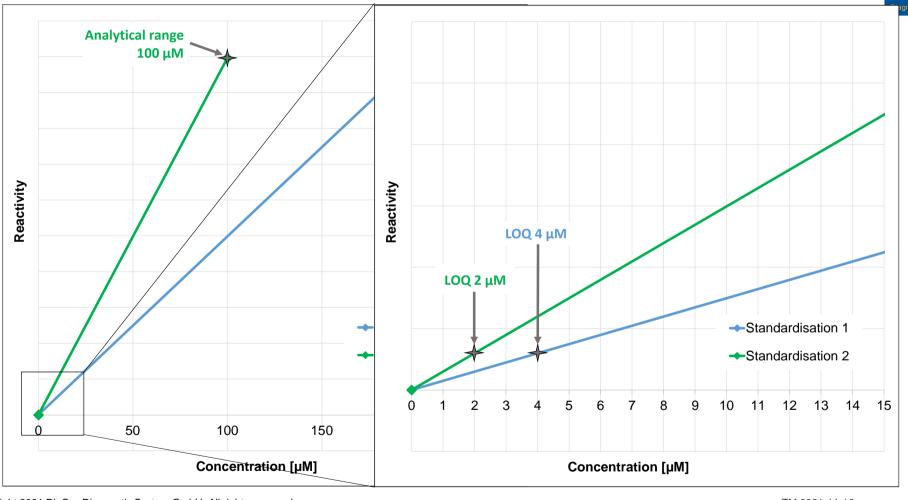
Annex I – Chapter II "Requirements regarding performance, design and manufacture"

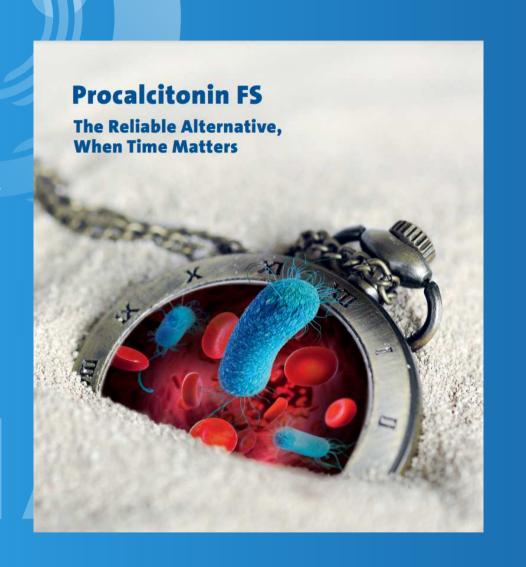
9.3 Where the performance of devices depends on the use of calibrators and/or control materials, the metrological traceability of values assigned to calibrators and/or control materials shall be assured through suitable reference measurement procedures and/or suitable reference materials of a higher metrological order. Where available, metrological traceability of values assigned to calibrators and control materials shall be assured to certified reference materials or reference measurement procedures.

Direct effects of a (missing) standardisation







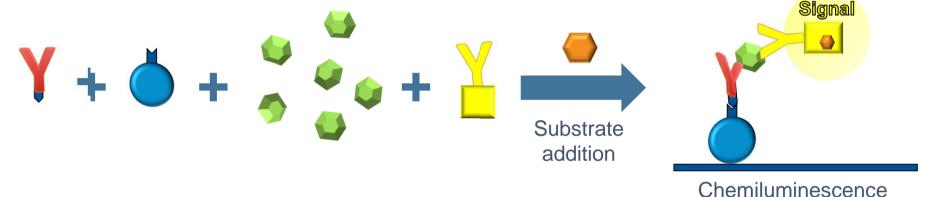


Procalcitonin



Chemiluminescence Immunoassay (CLIA) technology







Biotin-labelled antibody



Streptavidin labelled-magnetic beads



Antigen



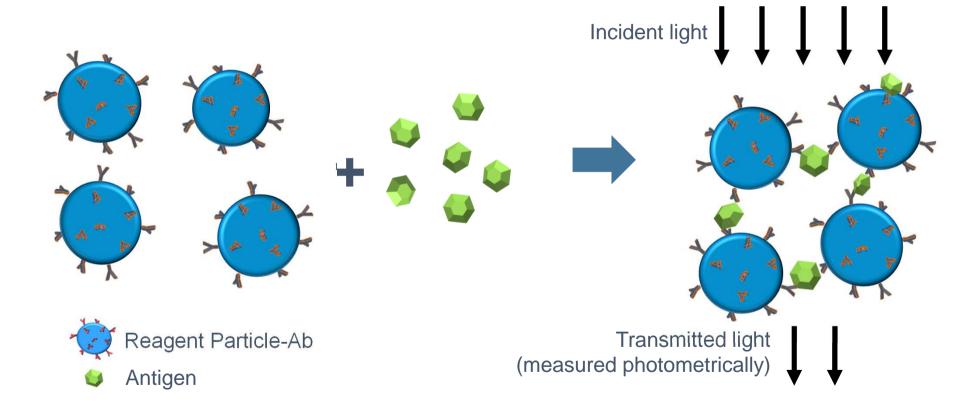
Detection system labelled-antibody (Acridinium-ester, Ruthenium, Luminol, etc.)



Substrate

Particle Enhanced Turbidimetric Immunoassay (PETIA) technology





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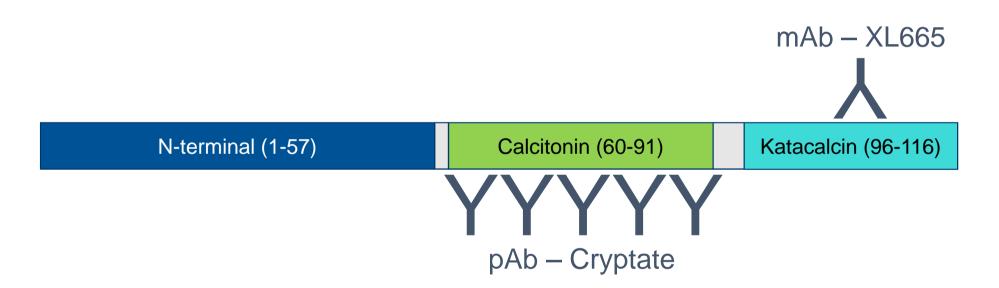
CLIA vs. PETIA – Example PCT



Parameter	CLIA	PETIA
Precision	++ (CV% ca. 2% @0.49 μg/L)	+ (CV% ca. 7% @0.45 μg/L)
Functional sensitivity (LOQ)	++ (0.06 µg/L ca.)	+ (0.2 µg/L ca.)
Analytical range (Δ linearity-LOQ)	++ (up to 100 μg/L)	+ (50 μg/L)
Interferences	++	+
Stability	++	++
Cost structure	High	Low
Sample volume	High (average ca. 100 µL)	Small (ca. 10-15 µL)
Waste production	Disposable materials	No disposable materials needed
Applicability	Low (instrument specific)	Very high (every photometric system)
Time-to-result	Longer (18-40 min)	Short (10 min)

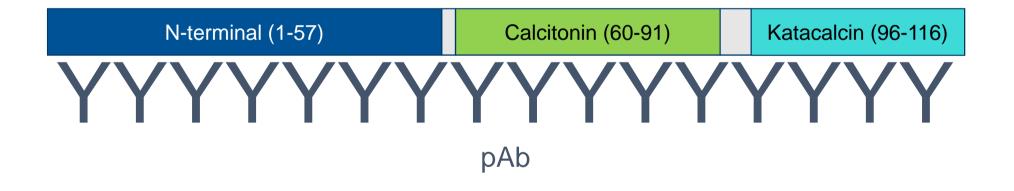


BRAHMS PCT Sensitive Kryptor





DiaSys PCT FS (PETIA)





Architect BRAHMS PCT (Abbott)

mAb (mouse) – Acridinium (tracer)

人(23-101)

N-terminal (1-57)

Calcitonin (60-91)

Katacalcin (96-116)

Y

mAb (rat) – Mag Beads (capture)



Elecsys BRAHMS PCT (Roche Diagnostics)

mAb (mouse) – Ruthenium (tracer)



N-terminal (1-57)

Calcitonin (60-91)

Katacalcin (96-116)



mAb (mouse) – Mag Beads (capture)

Huynh H. et al., 2021 https://doi.org/10.1016/j.cca.2021.01.004



Lumipulse G BRAHMS PCT (Fujirebio)

N-terminal (1-57)

Liaison BRAHMS PCT II Gen (Diasorin)

mAb – Alkaline phosphatase (tracer)
mAb – Mag Beads (capture)

Calcitonin (60-91)

Katacalcin (96-116)

mAb - Mag Beads (capture)

mAb – Isoluminol (tracer)



Calibrator standardization on an already existing immunoassay (used as reference)

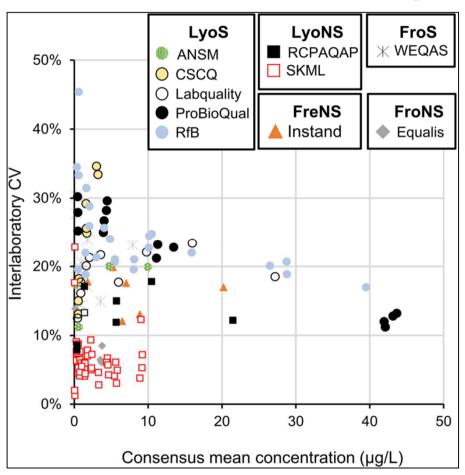


	Gravimetric value [ng/mL]	CLIA A [ng/mL]	CLIA B [ng/mL]
Level 5	53.52	26.01	27.10
Level 4	26.58	15.41	11.92
Level 3	12.65	7.74	5.60
Level 2	5.08	2.27	2.21
Level 1	1.15	0.46	0.54
Level 0	0.00	0.00	0.03

Uncertainty unknown

Harmonization through EQAs

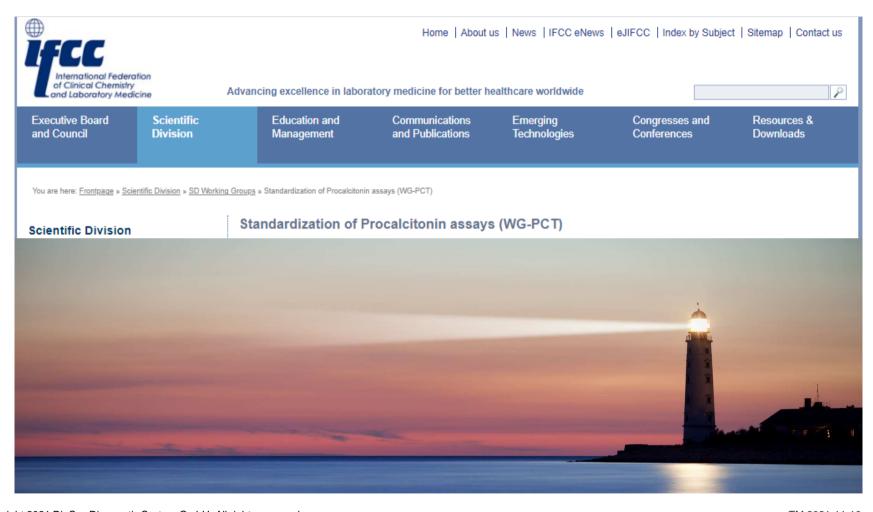


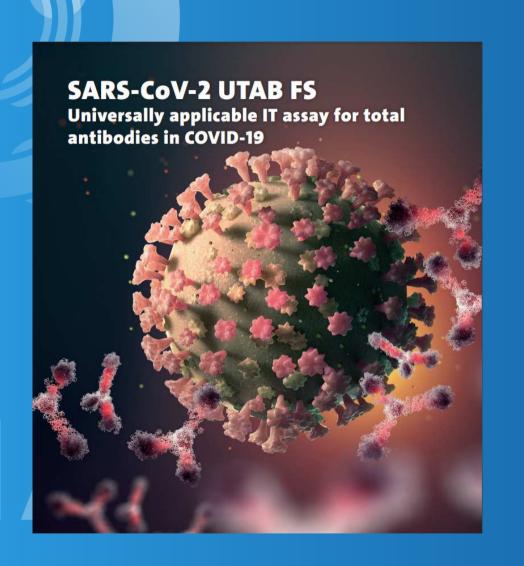


- Interlaboratory CVs are highly heterogeneous across the different surveys
- Materials with varying concentrations of PCT and various types of matrices (fresh, frozen, lyophilized, spiked with recombinant PCT or not) were used
- 3. Are these materials commutable?

IFCC PCT Standardisation Working Group







SARS-CoV-2







Medicines & Healthcare products Regulatory Agency

WHO International Standard
First WHO International Standard for anti-SARS-CoV-2
immunoglobulin (human)
NIBSC code: 20/136
Instructions for use
(Version 2.0, Dated 17/12/2020)

3. UNITAGE

The assigned potency of the WHO International Standard for SARS-CoV-2 is 250 IU/ampoule for neutralising antibody activity. After reconstitution in 0.25 mL of distilled water, the final concentration of the preparation is 1000 IU/mL.

For binding antibody assays, an arbitrary unitage of 1000 binding antibody units (BAU)/mL can be used to assist the comparison of assays detecting the same class of immunoglobulins with the same specificity (e.g. anti-RBD IgG, anti-N IgM, etc.)

Differences between WHO-standarised tests



Test 1

Test 3

Test 4

Test 5

v = 82.5 + 15.54 xTest 2 tAb [U/mL]

$$y = 33.4 + 2.18 \times \frac{200}{1000} = 33.4 + 2.1$$

$$y = 12.3 + 0.65 \text{ X}$$

$$y = 24.5 + 0.13$$
Test 1 IgG [AU/mL]

$$y = 34.5 + 0.02$$
Test 1 lgG [AU/mL]

$$y = 6.2 + 0.04 \text{ x}$$
Test 1 lgG [AU/mL]

$$y = 30.5 + 0.16$$

$$y = 30.5 + 0.16$$

$$0 = 30.5 + 0.16$$

$$0 = 30.5 + 0.16$$
Test 3 [AU]mL]

$$R = 0.94$$

$$y = 0.31 \times$$
0 200 400 600 800 1000

Test 3 [AU/mL]

Perkmann et al., 2021

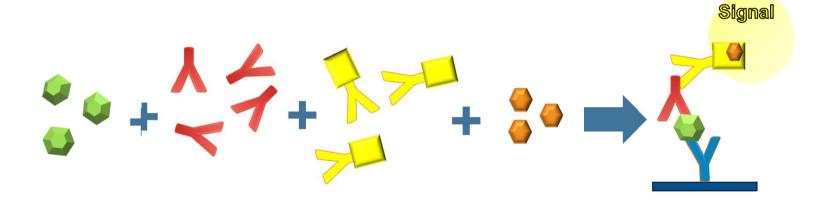
Anti-Spike Protein Assays to Determine SARS-CoV-2 Antibody Levels: a Head-to-Head Comparison of Five Quantitative Assays https://doi.org/10.1128/Spectrum.00247-21

v = -50.9 + 1.78 x100 150 200 TM 2021-11-10 Test 4 [AU/mL]

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Enzyme-linked immunosorbent assay (ELISA)







- Y Plate immobilised antibody
- Antigen (SARS-CoV-2 protein)
- Detection antibody, bound to POD
- Substrate (TMB)



Additional efforts shall also be made for improving harmonization, standardization and comparability of anti-SARS-CoV-2 serology.

Plebani et al., 2021

SARS-CoV-2 antibody assay after vaccination: one size does not fit all https://doi.org/10.1515/cclm-2021-0703



Calprotectin

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Non-homogeneity of measurement procedures



Greatest problem is the matrix itself: stool

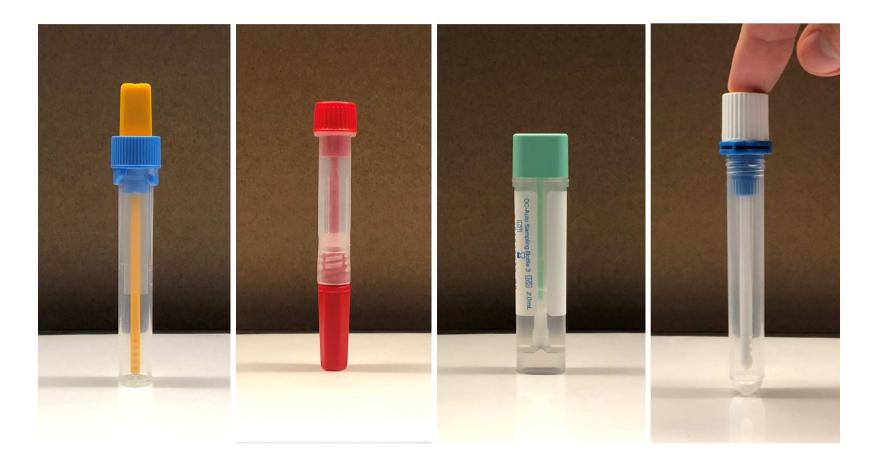
- Stool is highly inhomogene in comparison to serum / plasma
- Interferences by host proteins, salts / ions, bacterial proteins, lipids etc.
- Process of dissolution in specific matrix tedious and not standardized

Stool sample tubes for clinical analyzers

- Differences by manufactures in sizes, forms, amount of stool and collecting systems
- Amount of stool collected reproducible?
- Handling of stool samples varies with tube (e.g. vortex), preparation and dilution factor

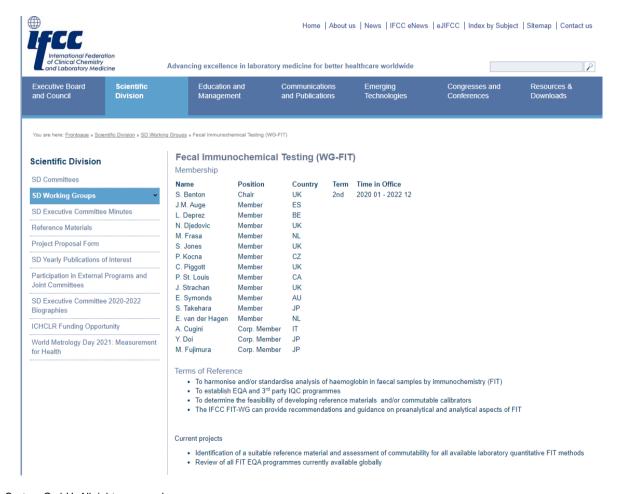
Influence of the stool tubes



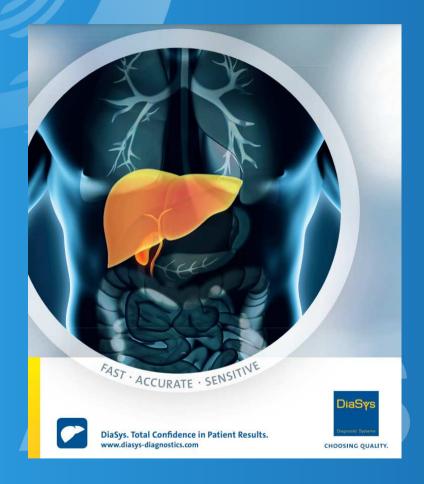


IFCC Fecal Immunochemical Testing (WG-





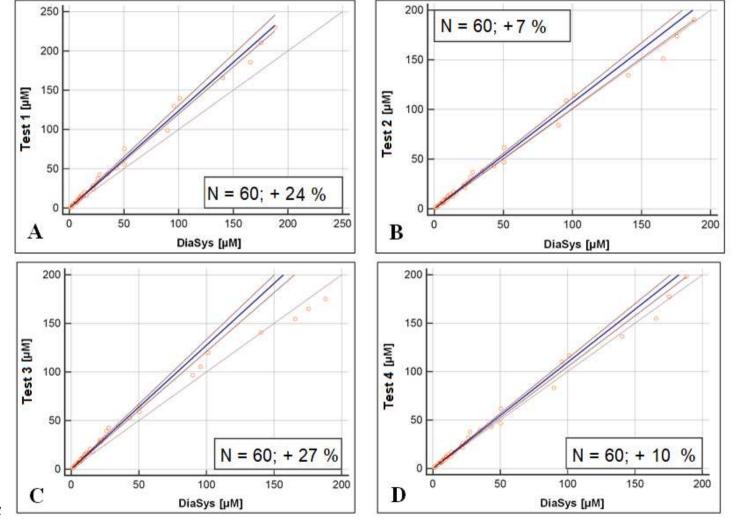
Total Bile Acids 21 FS





DiaSys vs. commercial tests P&B regression



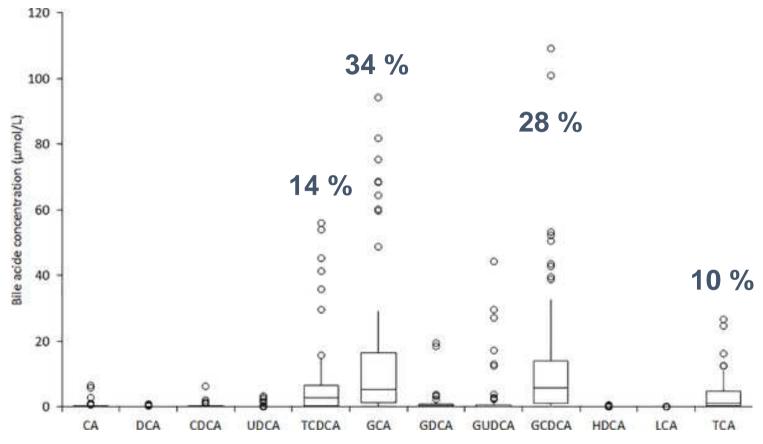


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21-11-10

Bile acids composition of plasma samples



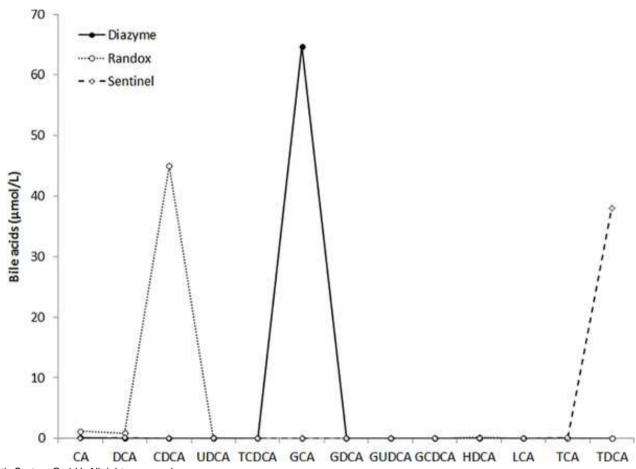


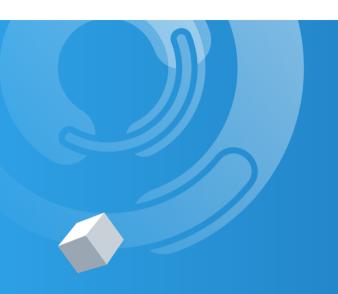
Danese et al., 2017

Analytical evaluation of three enzymatic assays for measuring total bile acids in plasma using a fully-automated clinical chemistry platform https://doi.org/10.1371/journal.pone.0179200

Bile acids composition of calibrators of three commercial assays







Futher parameters

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The long and winding road...



Parameter	Standardisation / measurement issues
β-Hydroxybutyrate	No reference material/method; different Enzymes used by manufacturer + D/L impurity of calibration material
LpPLA2	No reference material; Enzyme activity vs. mass
ACE	No reference material, method; different matrices; enzyme vs. mass
Cys-C	No commutable reference material on diverse matrices (serum / urine)
Presepsin	No reference material, method
D-Dimer	No reference material, method; update
APOs	Update of reference material, method

Conclusions



- Regulations and guidelines, e.g. the new IVDR 2017/746 (EU), strengthen the traceability aspects "through suitable reference measurement procedures and/or suitable reference materials of a higher metrological order"
- Standardisation (recovery) issues do affect assay parameters (e.g. LOQ) and development process (e.g. reaction enhancer, beads concentrations, etc.)
- Different conditions (mAb-pAntibodies, epitopes, labelling, detection systems, techniques) or enzymes can lead to different quantitations → importance of the specificity
- Absence of reference material (CRM) implies harmonization / standardisation of results through different methods (EQA, etc.)
- Defining the sample matrix (blood, stool, liquor, urine) and the commutability



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